## Adoptive Immunotherapy

- Greatest initial success was in the eradication of relapsed CML after allogeneic transplant.
- Also useful in treatment of post-transplant EBVlymphoproliferative disease
- Initial use of unselected leukocytes
- Dosed according to content of CD3+ T cells
- Other component leukocytes, e.g., NK cells, very likely play a role.

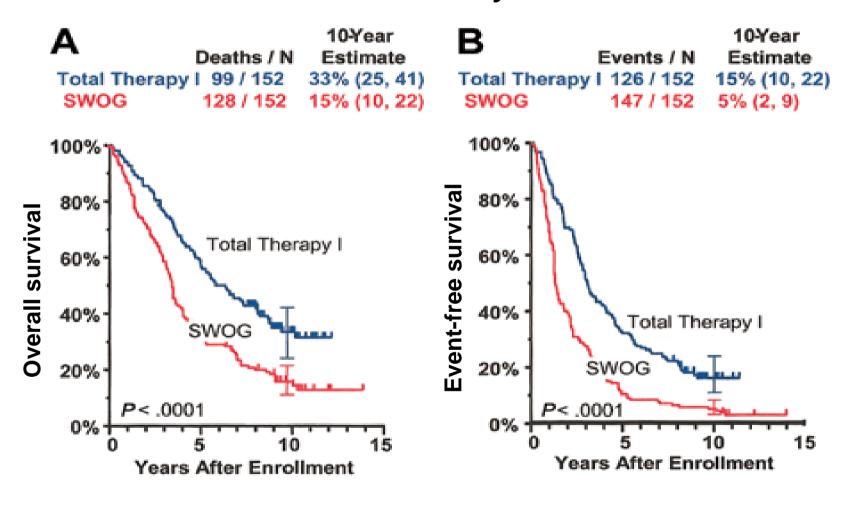
## Adoptive Immunotherapy

- Success of donor leukocyte infusions has led to the incorporation of immunotherapy as an integral feature of preparative regimens.
- Nonmyeloablative preparative regimens shift the burden of tumor eradication from high-dose conditioning regimens to the donor's immune cells

## Adoptive immunotherapy

- Targeted therapy to enhance anti-tumor or antipathogen effect without exacerbating GvHD
  - tumor or virus specific Ag
  - minor histocompatibility Ags
  - specificity conferred by the APC, e.g., DCs
- Role of cytokines in expansion ex vivo or adjuvant administration in vitro.
- Route of administration

Comparison of standard chemoRx (SWOG) vs chemoRx→autograft→IFN maintenance (Total Therapy I), median f/u 9yrs



Barlogie et al., <u>Blood</u>, <u>103</u>: 20-32, 2004

## Clinical protocols using dendritic cells to generate Ag-specific T cells for adoptive immunotherapy

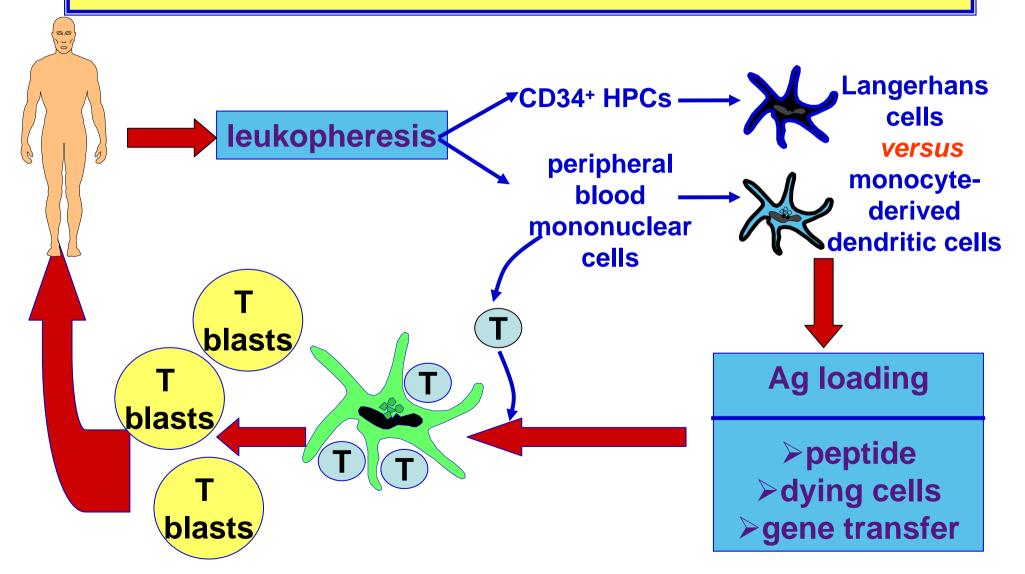


Table 1. Different Sources of Antigens and Antigen-Presenting Cells in Preclinical and Clinical Studies Generating CMV-Specific CTLs

| Antigen                                                | Antigen-Presenting Cell                        | Reference                 |
|--------------------------------------------------------|------------------------------------------------|---------------------------|
| Virions                                                | Skin fibroblast                                | Walter et al. [51]        |
| Retroviral vector encoding pp65                        | B lymphoblastoid cell line                     | Sun et al. [36]           |
| Adenoviral vector encoding pp65                        | Dendritic cells                                | Keever-Taylor et al. [56] |
|                                                        |                                                | Hamel et al. [72]         |
| Adenoviral vector encoding pp65                        | Dendritic cells and B lymphoblastoid cell line | Sifi et al. [58]          |
| CMV antigen derived from CMV-infected lung fibroblasts | Dendritic cells                                | Peggs et al. [54]         |
| CMV lysate and antigen                                 | Peripheral blood mononuclear cells             | Einsele et al. [55]       |
| HLA-A*0201-restricted CMV peptide pp65(495–503)        | Dendritic cells and B lymphoblastoid cell line | Szmania et al. [92]       |
| HLA-A*0201-restricted CMV peptide pp65(495–503)        | Dendritic cells                                | Foster et al. [93]        |

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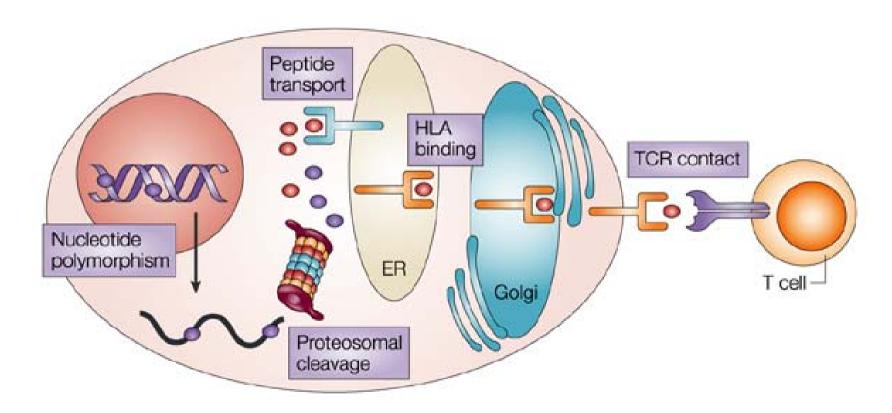
Table 2. Published Reports on Use of EBV-Cytotoxic T Cells as Prophylaxis or Treatment for PTLD after BMT

| Study                                         | No.<br>Pts.<br>(Age)    | Type of Transplant                                                                                                                                    | Path.<br>Evidence<br>of PTLD                                   | Cytotoxic T-Cell<br>(CTL) Lines and<br>Dose                                                                                                     | Results                                                                                                                                                         |
|-----------------------------------------------|-------------------------|-------------------------------------------------------------------------------------------------------------------------------------------------------|----------------------------------------------------------------|-------------------------------------------------------------------------------------------------------------------------------------------------|-----------------------------------------------------------------------------------------------------------------------------------------------------------------|
| Rooney et                                     | 39 (9<br>mo to<br>20 y) | T cell-depleted HSCT<br>(mismatch related donor or<br>matched unrelated donor)                                                                        | No—<br>prophylax<br>is study                                   | Allogeneic (donorderived) EBV CTL: minimum dose of 4 x 10 <sup>7</sup> /m <sup>2</sup> and maximum dose of 12 x 10 <sup>7</sup> /m <sup>2</sup> | No patients developed PTLD compared with 11.5% of controls: no toxicity                                                                                         |
| Rooney et al. [32] and Gottschalk et al. [33] | 3<br>(12–<br>17 y)      | T cell-depleted HSCT                                                                                                                                  | Yes—<br>lymphobl<br>astic<br>lymphom<br>a                      | Allogeneic (donor-<br>derived) EBV CTL<br>2–4 × 10 <sup>7</sup> /m <sup>2</sup>                                                                 | 2 compete remissions, 1 died (no response to CTL secondary to tumor mutation resistant to CTL)                                                                  |
| Gustafsson<br>et al. [13]                     | 6 (1–<br>39 y)          | T cell-depleted HSCT or<br>unmanipulated HSCT with<br>ATG/OKT3 conditioning<br>(mismatched or matched<br>unrelated donor or matched<br>related donor) | No—<br>treatment<br>based on<br>increased<br>EBV DNA<br>levels | Allogeneic (donor-<br>derived) EBV CTL 4<br>× 10 <sup>7</sup> /m <sup>2</sup>                                                                   | 5 patients had decreased EBV<br>DNA levels. 1 patient<br>subsequently died of PTLD<br>(CTL showed poor specificity<br>for EBV targets on cytotoxicity<br>assay) |

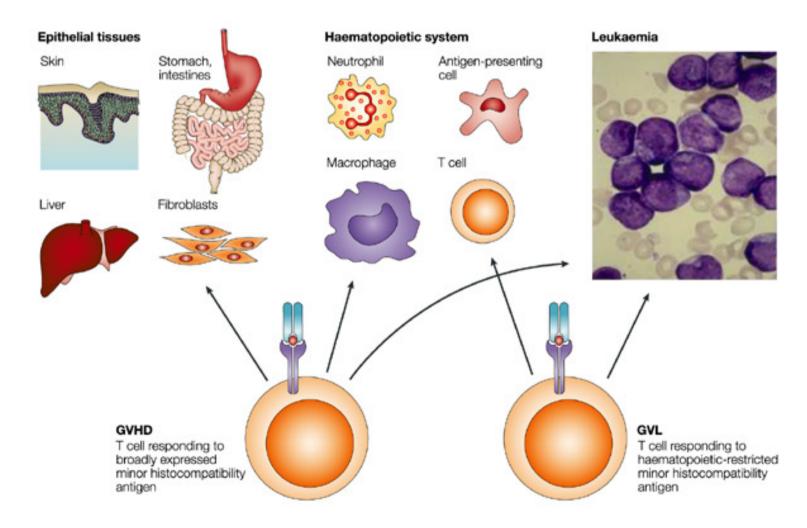
PTLD indicates posttransplantation lymphoproliferative disorder.

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# Generation of minor histocompatibility antigens



## Distinguishing GvL from GvHD



## Minor histocompatibility antigens

| Table 2   Advantages and disadvantages of candidate antigens |                                                                                                                                |                                                                                                        |  |  |  |
|--------------------------------------------------------------|--------------------------------------------------------------------------------------------------------------------------------|--------------------------------------------------------------------------------------------------------|--|--|--|
| Class of target                                              | Advantages                                                                                                                     | Disadvantages                                                                                          |  |  |  |
| Minor histocompatibility antigens                            | T cells have high avidity for<br>antigen; both CD8+ and CD4+<br>T cells recognize antigen;<br>potentially multivalent response | Limited to allogeneic<br>transplantation; limited<br>number of defined<br>antigens; potential for GVHD |  |  |  |
| Overexpressed normal proteins                                | Broad applicability for different types of cancer                                                                              | T cells have low avidity<br>for antigen; potential<br>for toxicity to normal<br>tissues                |  |  |  |

GVHD, graft-versus-host disease.

#### Adoptive immunotherapy with donor T cells to augment GvL

