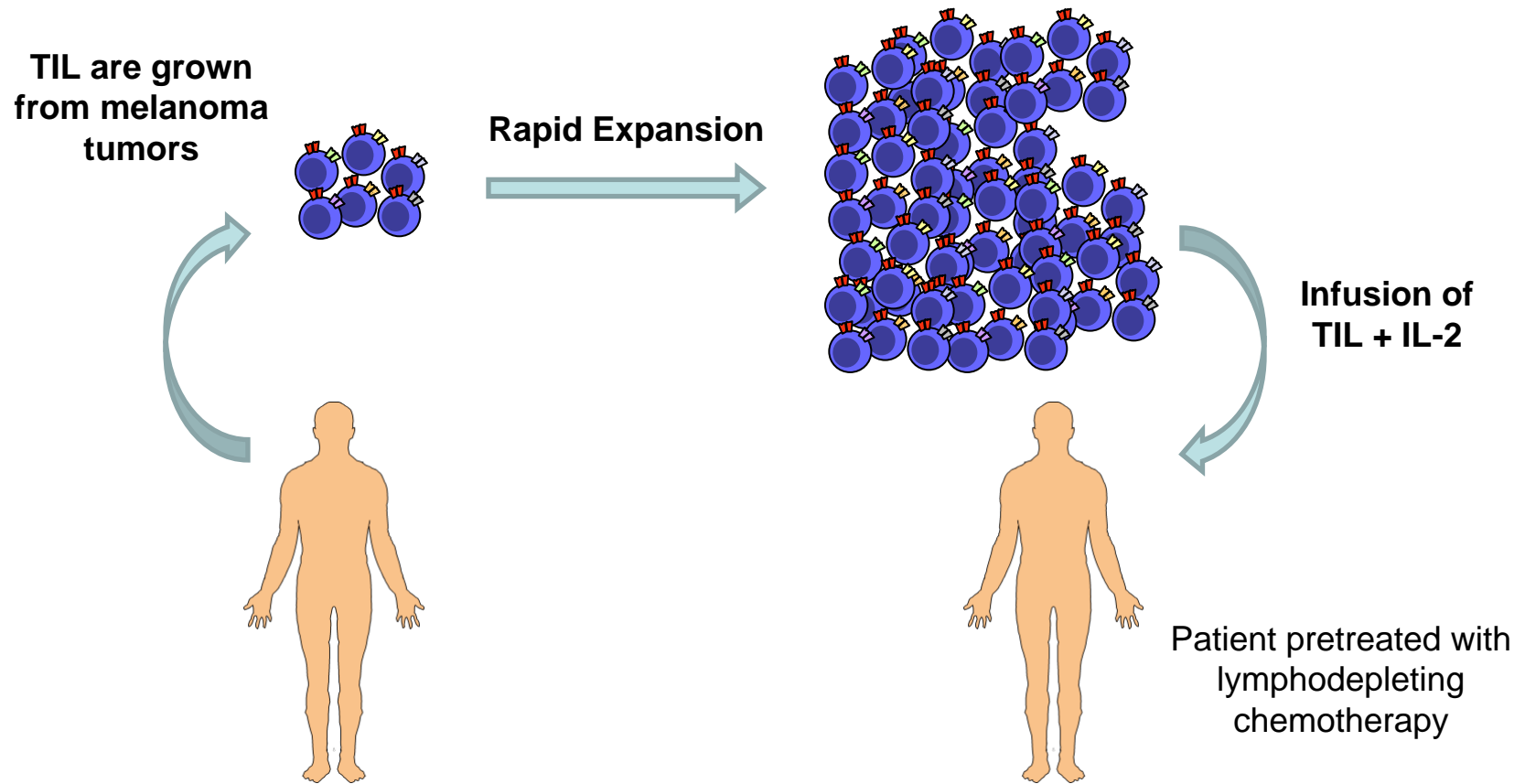


# Dissecting therapy-induced T-cell responses in melanoma

# Tumor-infiltrating lymphocyte (TIL) therapy of melanoma



- 50% response rate in trials in multiple centers (US, Israel)
- Clinical effect at least partially mediated by cytotoxic T cells



**Day -25**

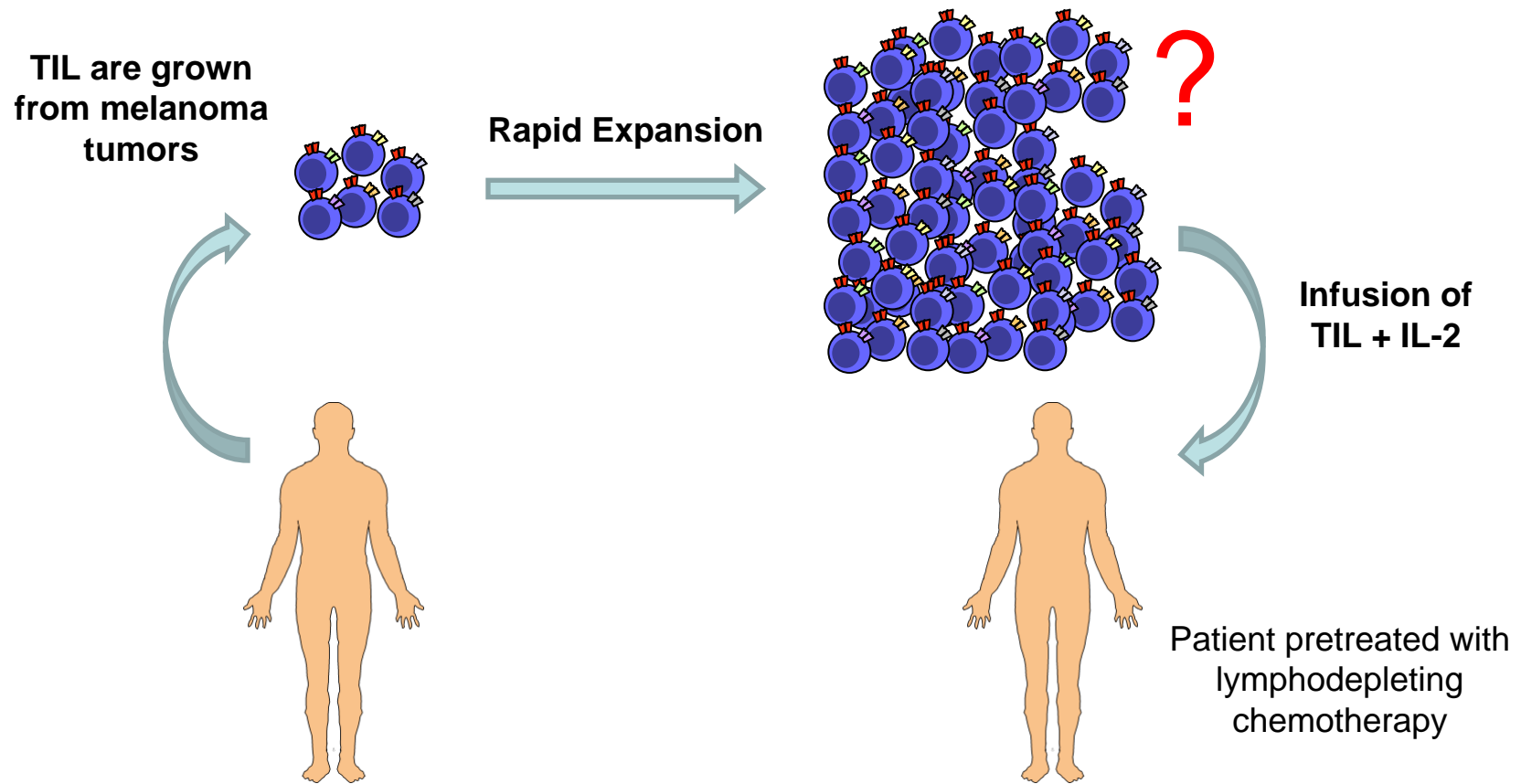


**Day +34**



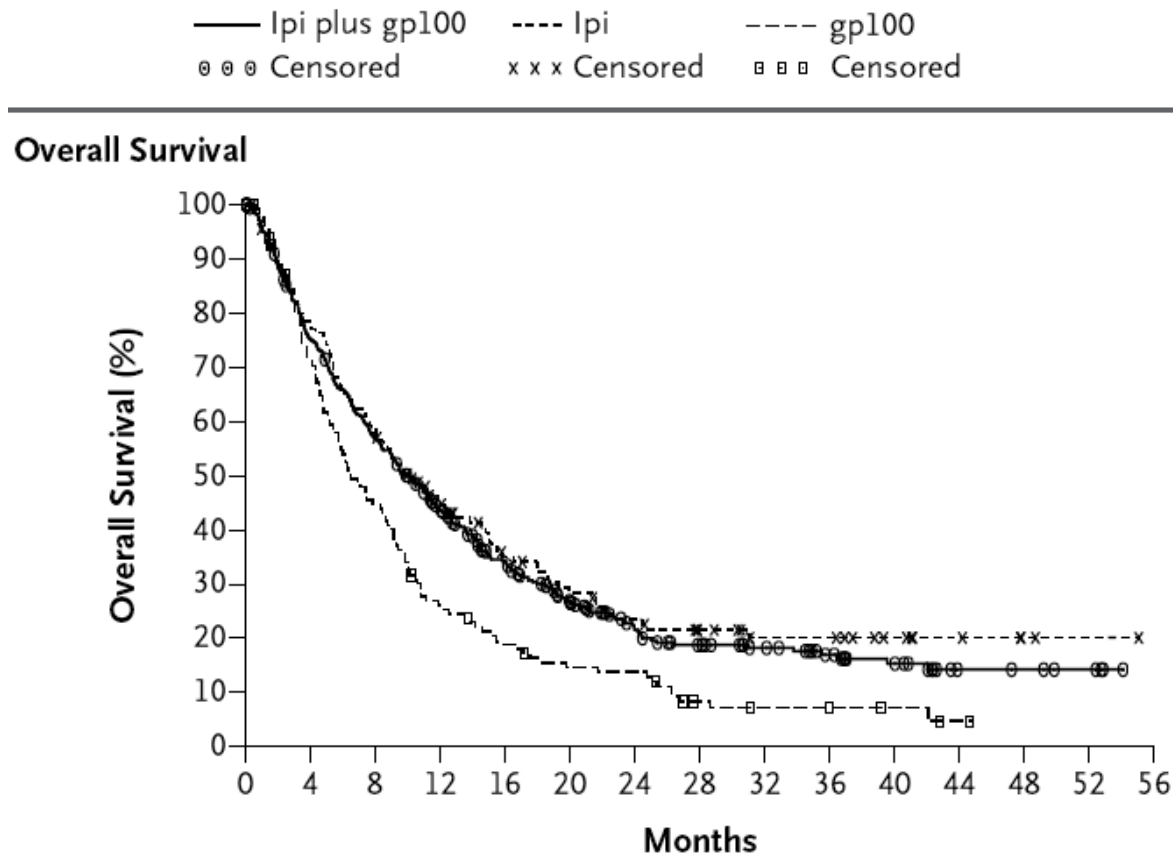
**3.2+ Years**

# Tumor-infiltrating lymphocyte (TIL) therapy of melanoma



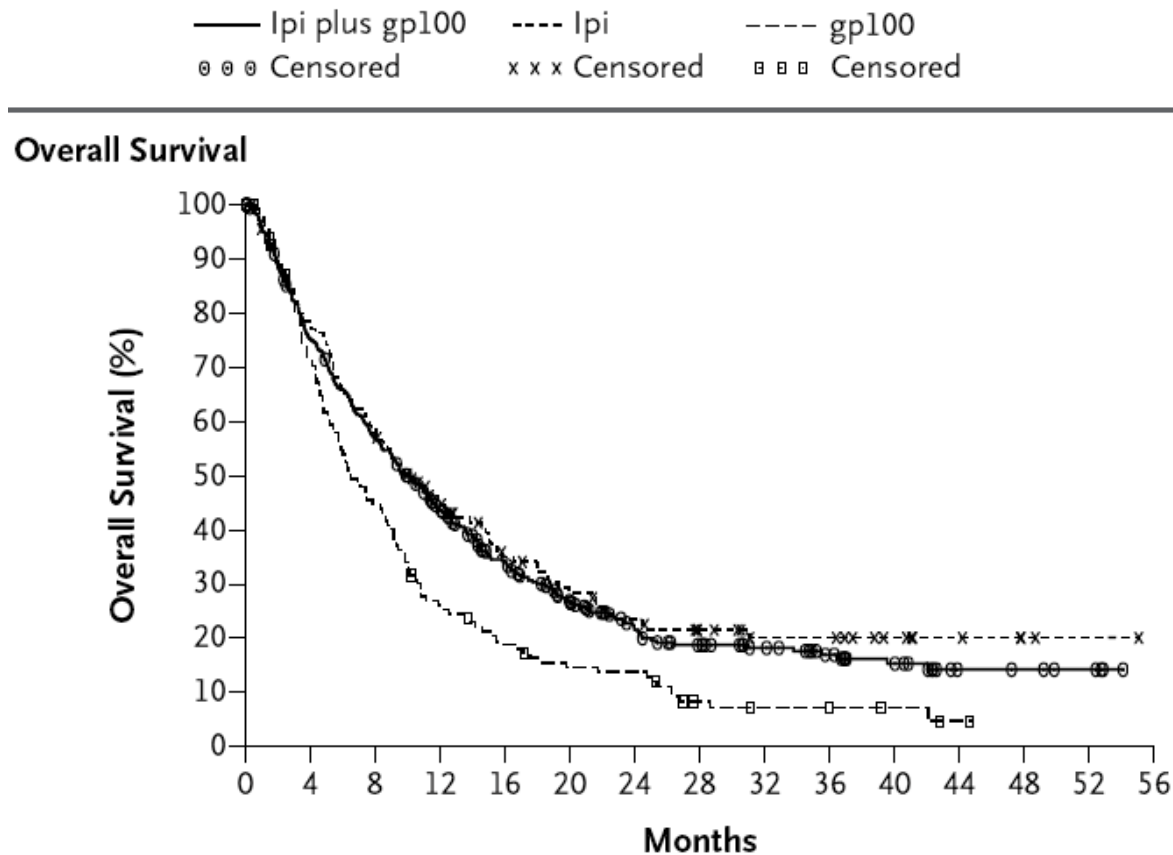
- Which cytotoxic T cells mediate cancer regression?
- Could we specifically boost their numbers?

# Anti-CTLA4 therapy of melanoma



- Survival benefit in metastatic disease
- Data from murine models suggest a role for cytotoxic T cells

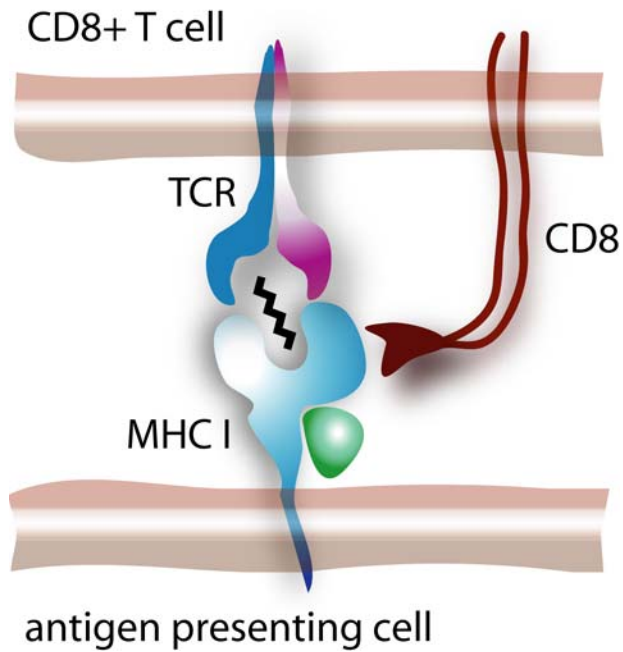
# Anti-CTLA4 therapy of melanoma



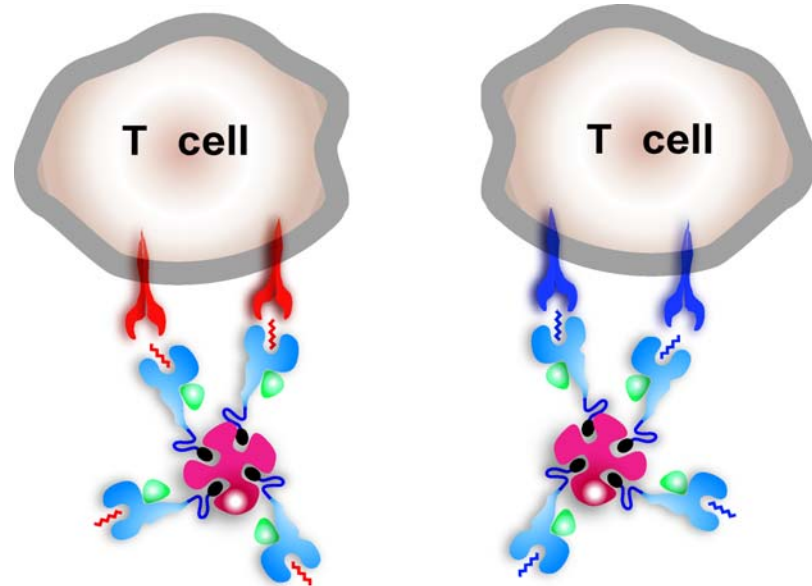
- Which cytotoxic T cells mediate cancer regression?
  - Could we specifically boost their numbers?
- Could we distinguish responders and non-responders?

## Detecting tumor-specific cytotoxic T cells

How a T cell sees a target cell

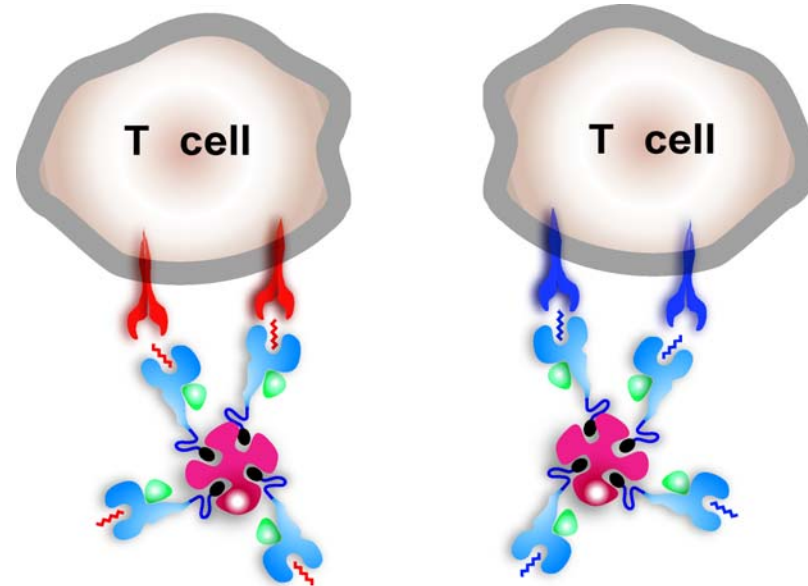
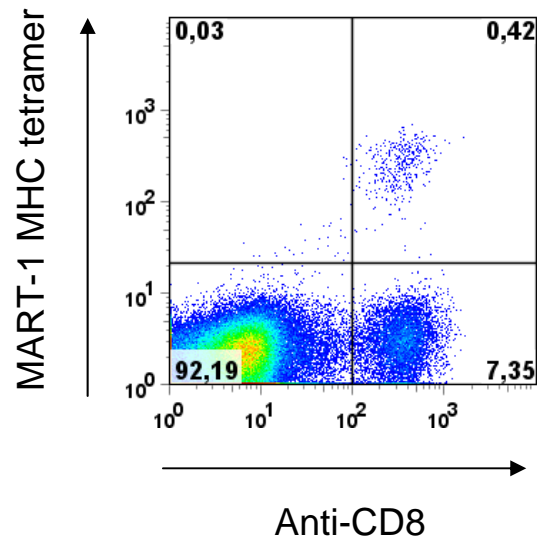


How an MHC tetramer sees a T cell



# Detecting tumor-specific cytotoxic T cells

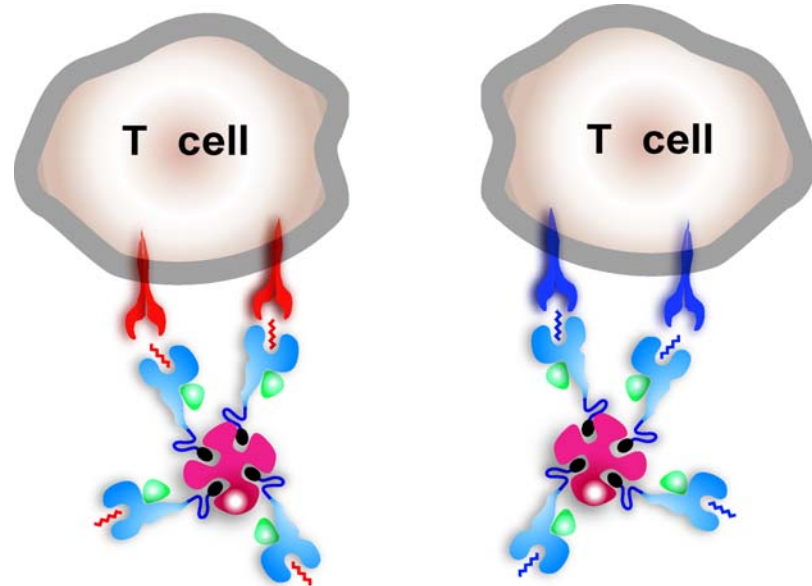
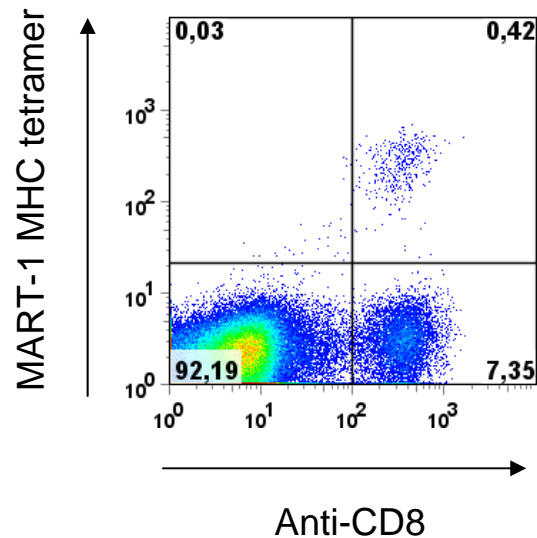
How an MHC tetramer sees a T cell





## Detecting tumor-specific cytotoxic T cells

How an MHC tetramer sees a T cell

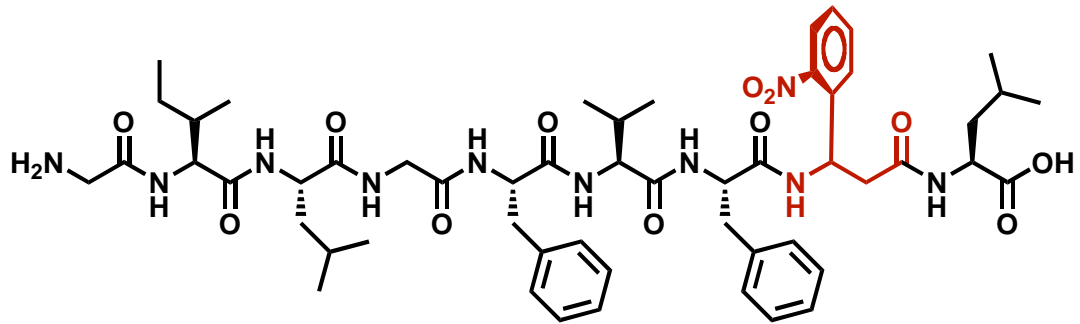


**Hundreds of different melanoma-associated T cell antigens have been described**

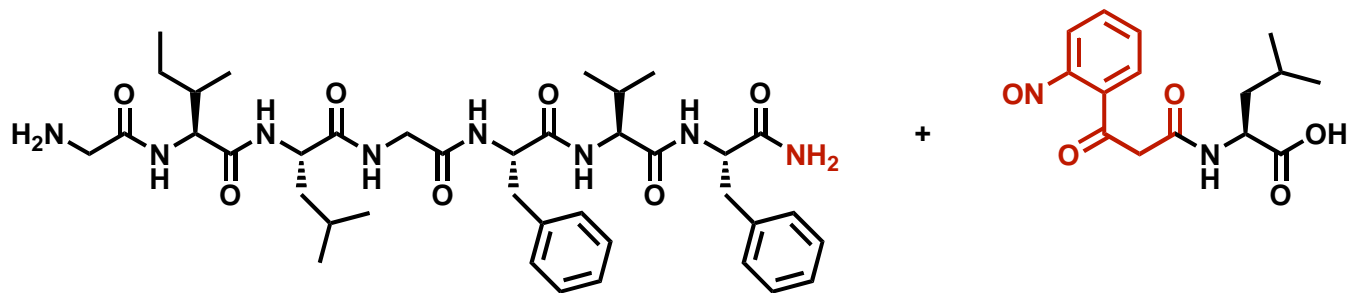
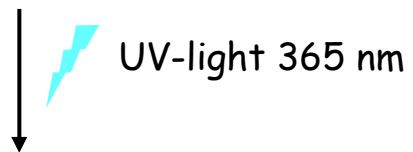
Two problems:

1. Generation of very large collections of MHC tetramer reagents is not possible with standard technology
2. Even if you would have such a collection, the amount of biological material is limiting

# Generation of pMHC multimers by UV-induced peptide exchange

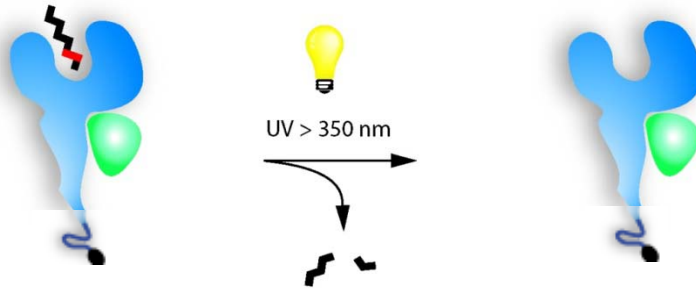


GILGFVF(*o*-NO<sub>2</sub>)L

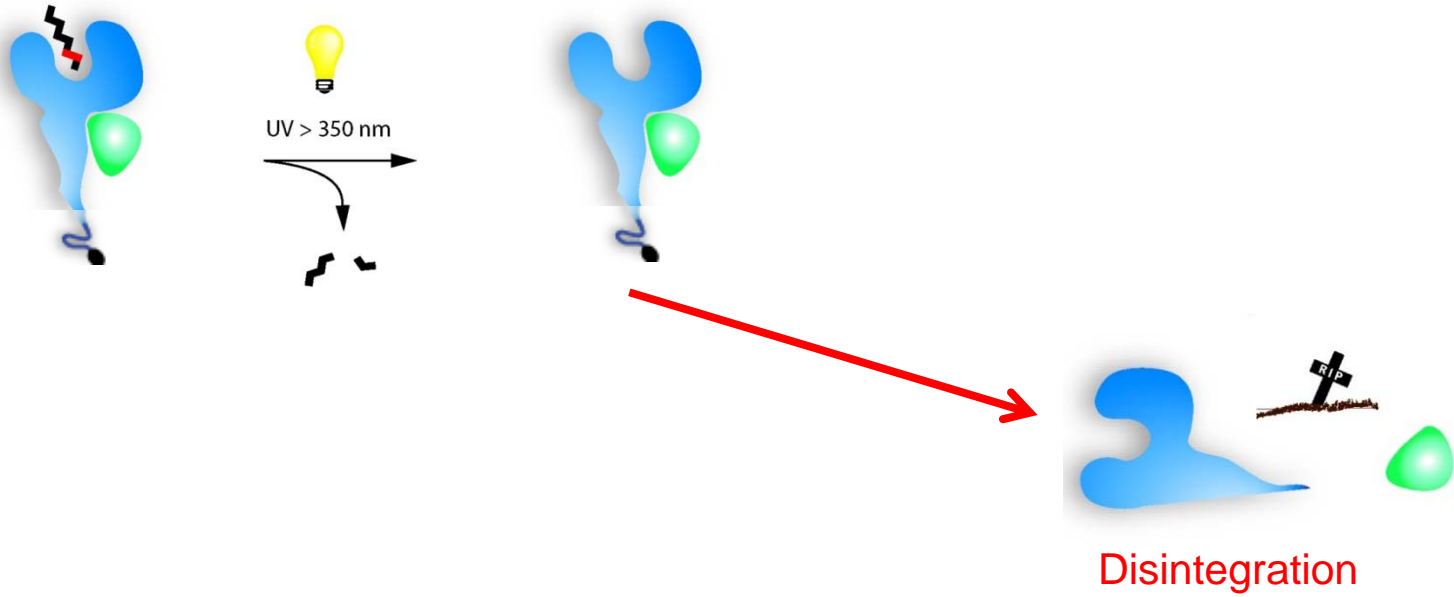


# Generation of pMHC multimers by UV-induced peptide exchange

---



# Generation of pMHC multimers by UV-induced peptide exchange





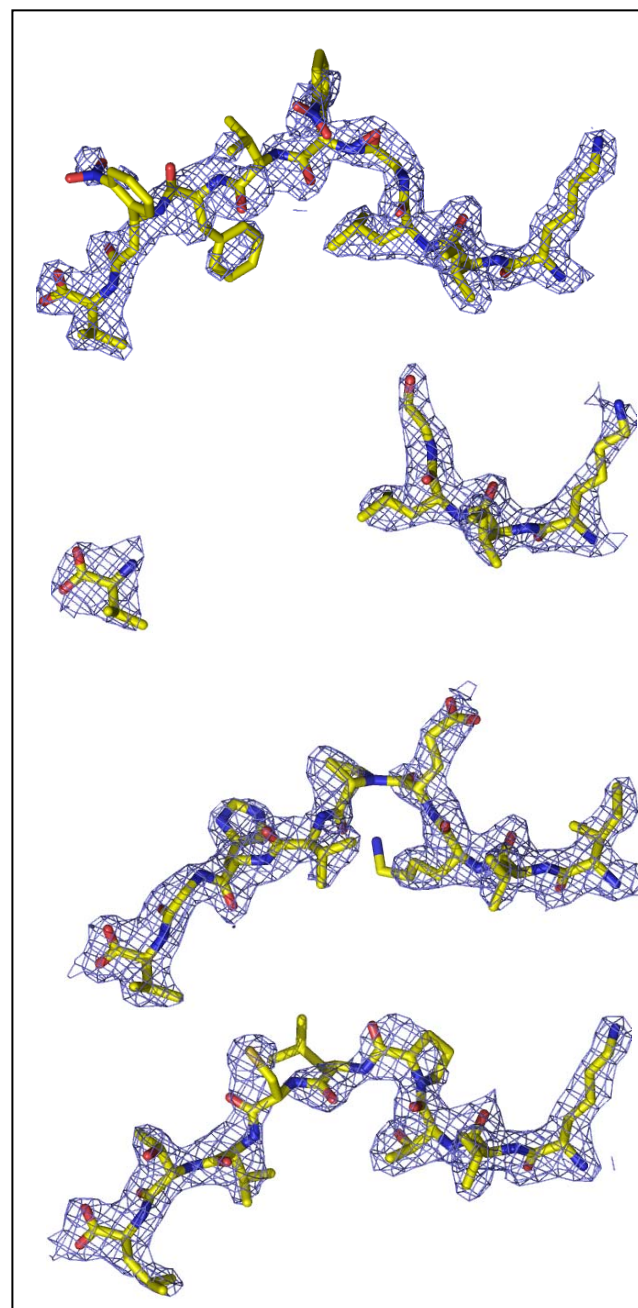
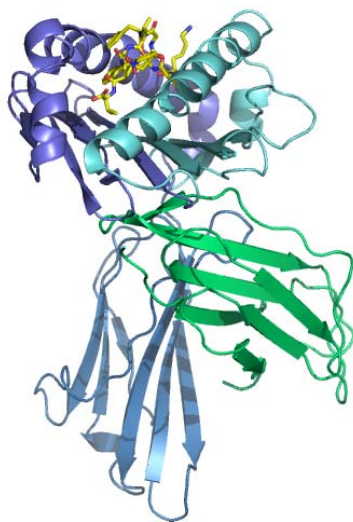
## *In crystallo* MHC ligand exchange

UV-sensitive ligand pre-cleavage  
(KILGJ\*VFJV)

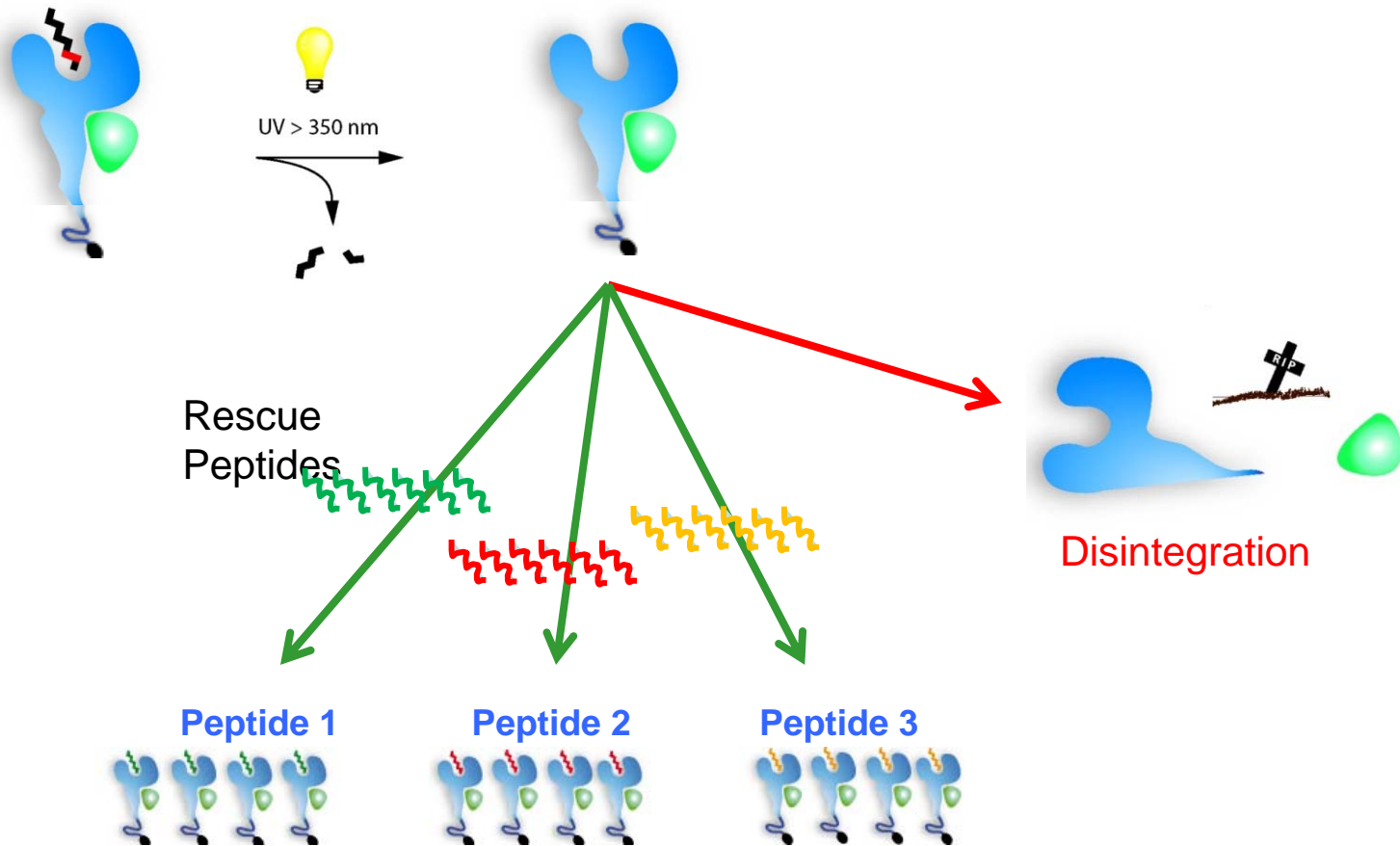
UV-sensitive ligand post-cleavage  
(visualization of the reaction intermediate)

*In crystallo* exchange with HIV RT<sub>468-476</sub>  
(ILKEPVHGV)

*In crystallo* exchange with HIV Env<sub>120-128</sub>  
(KLTPLCVTL)



# Generation of pMHC multimers by UV-induced peptide exchange



Allows the creation of very large MHC multimer collections for all prevalent HLA class I alleles

- HLA-A1, -A2, -A3, -A11, -B7, -B57 (Toebes et al. *Nat. Med.* 2006, Bakker et al. *PNAS* 2008)
- HLA-A24, -B40, -B58 (G. Grotenbreg, NUS, Singapore)

## How to monitor hundreds of T cell populations per patient?

---



Aim: Detection of low frequency T cell populations

~ 0.02% of CD8 T cells

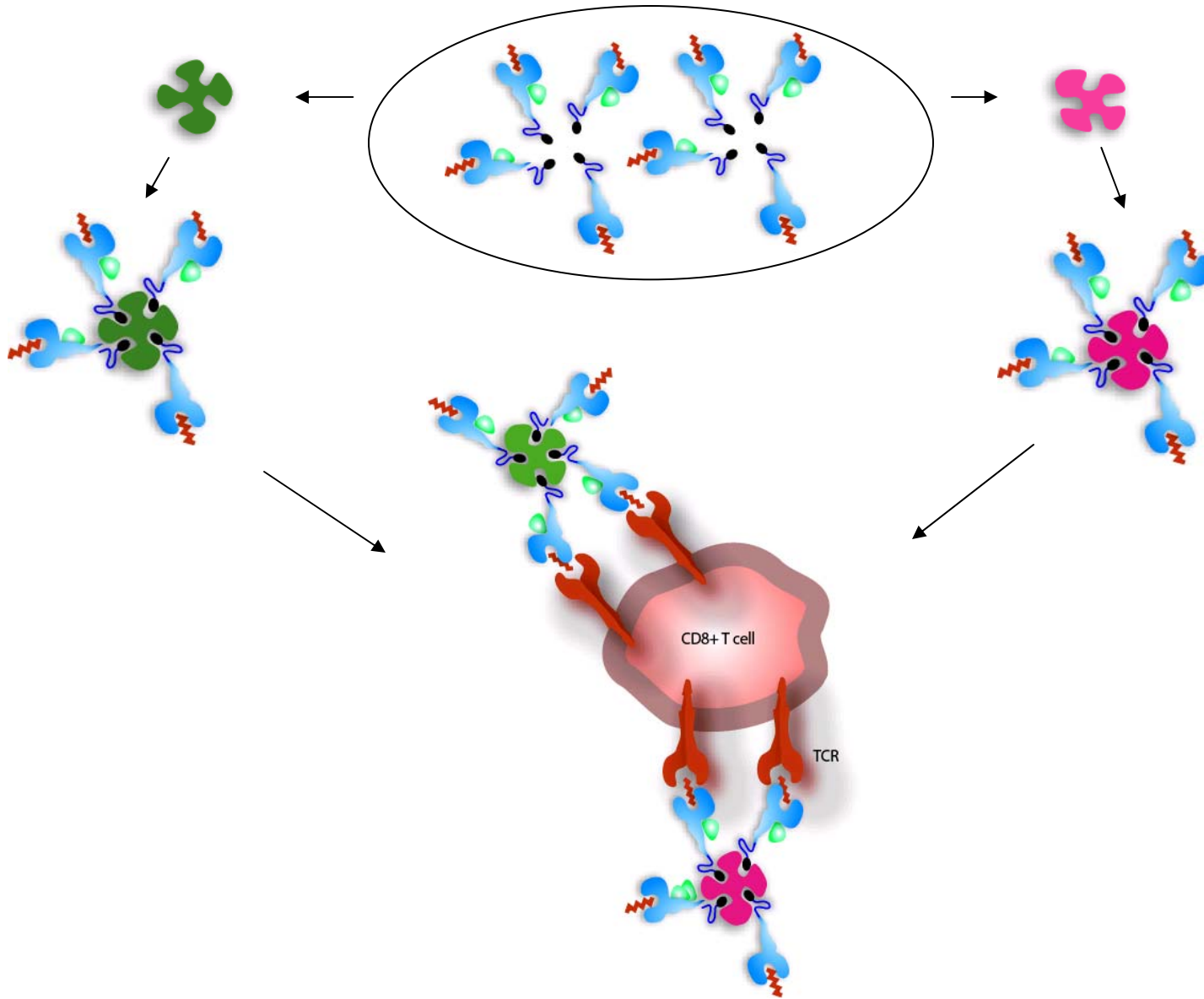
Required: record >50,000 CD8 T cells ~ 2 ml fresh blood

For 100 potential antigens: ~ 200 ml blood

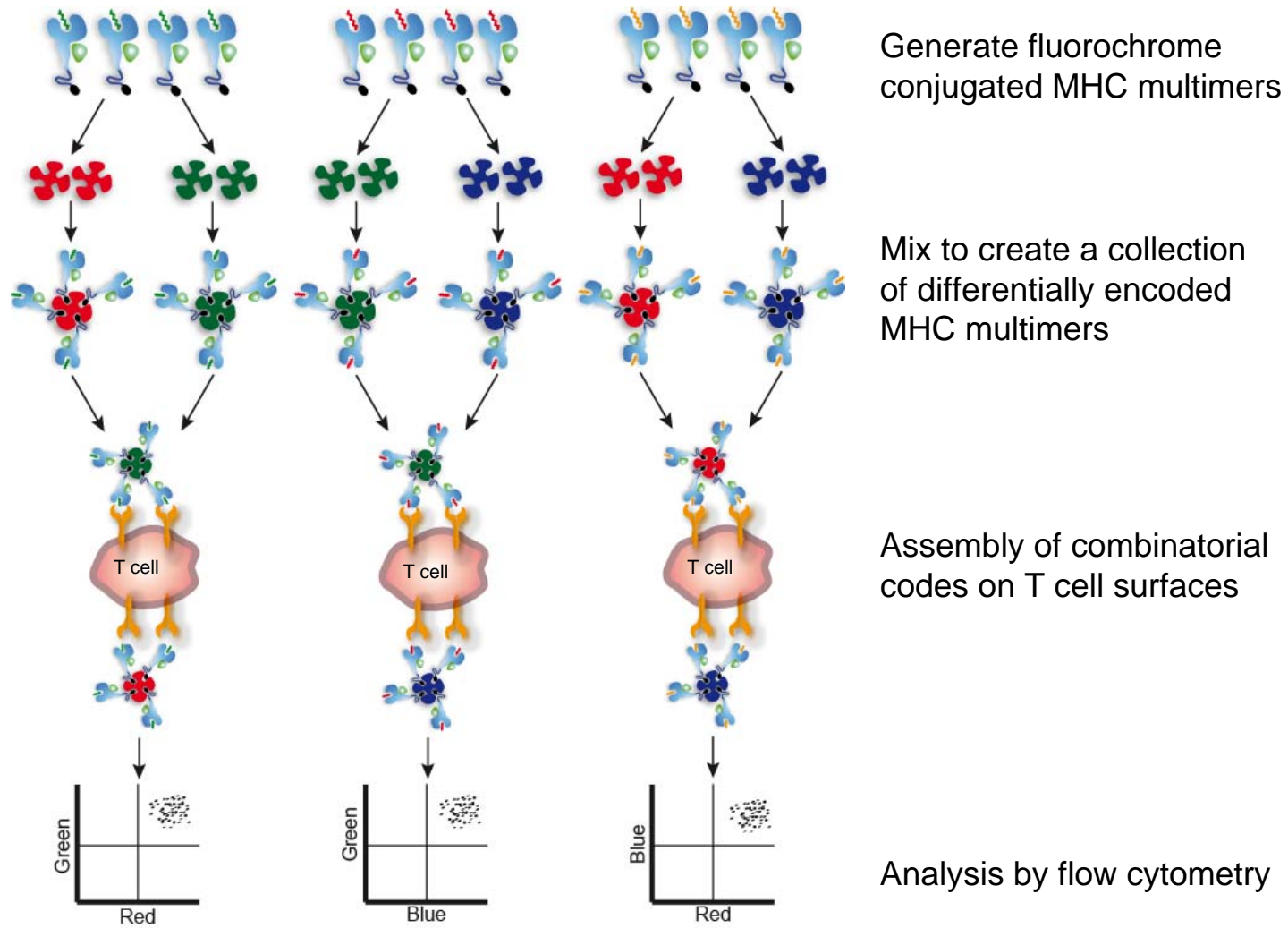
25 parallel measurements per sample: ~ 8 ml blood



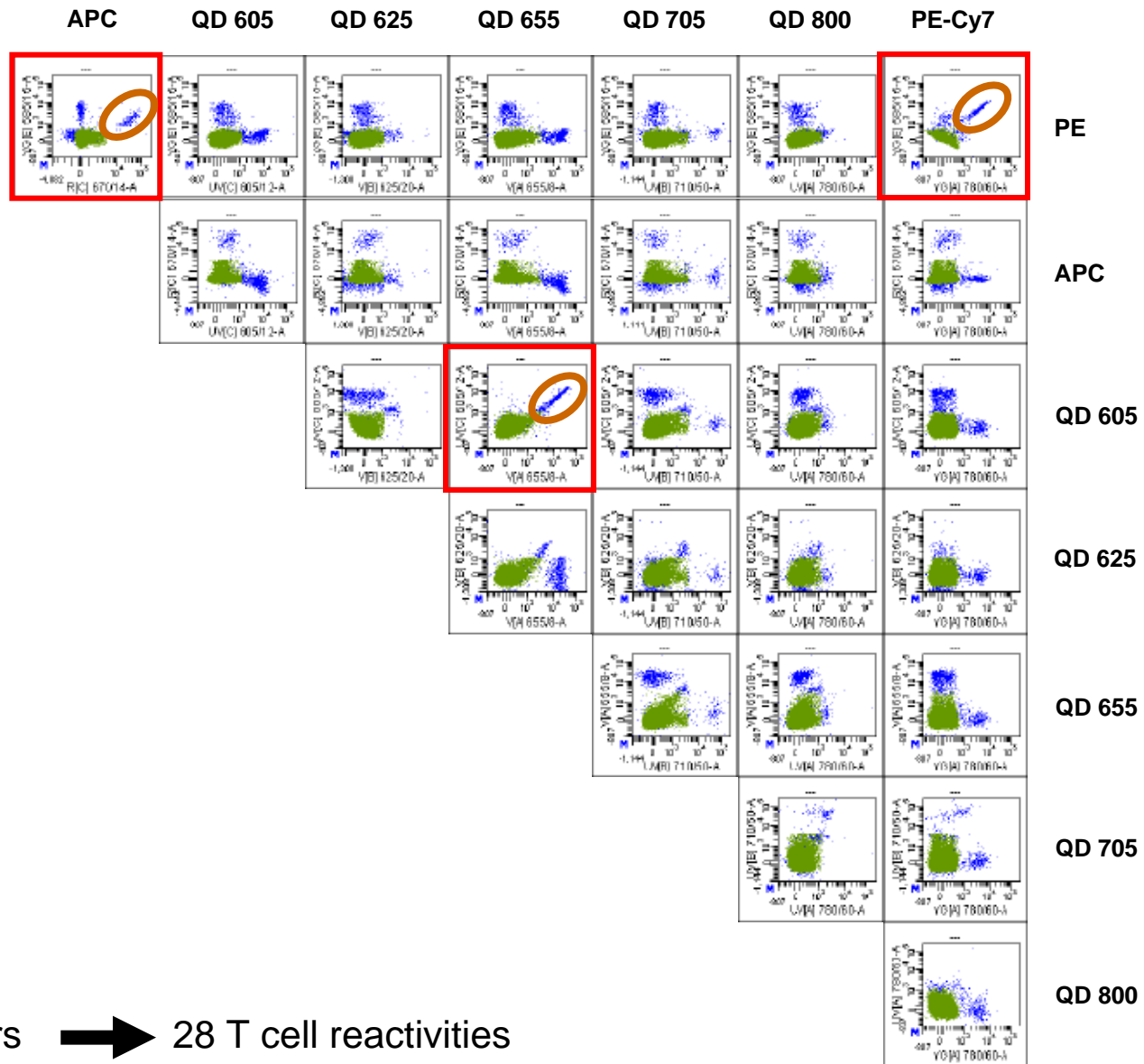
# What if you couple the same pMHC to multiple fluorophores & start combining?



# Self-assembling molecular codes



# Example of a combinatorial coding read-out



8 colors → 28 T cell reactivities

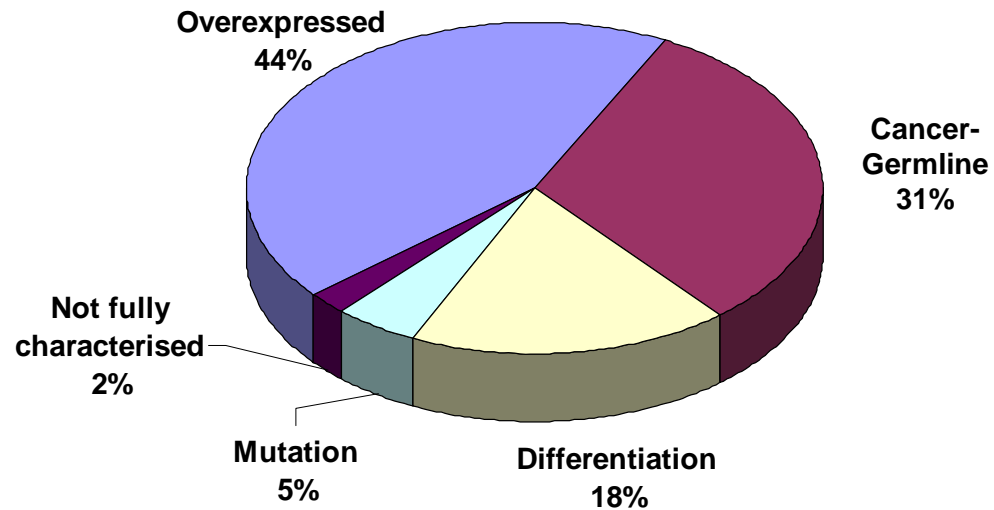
## Selection of melanoma-associated epitope panel

---

Panel of 216 melanoma-associated peptides

- public databases
- literature analysis

### Antigen class distribution

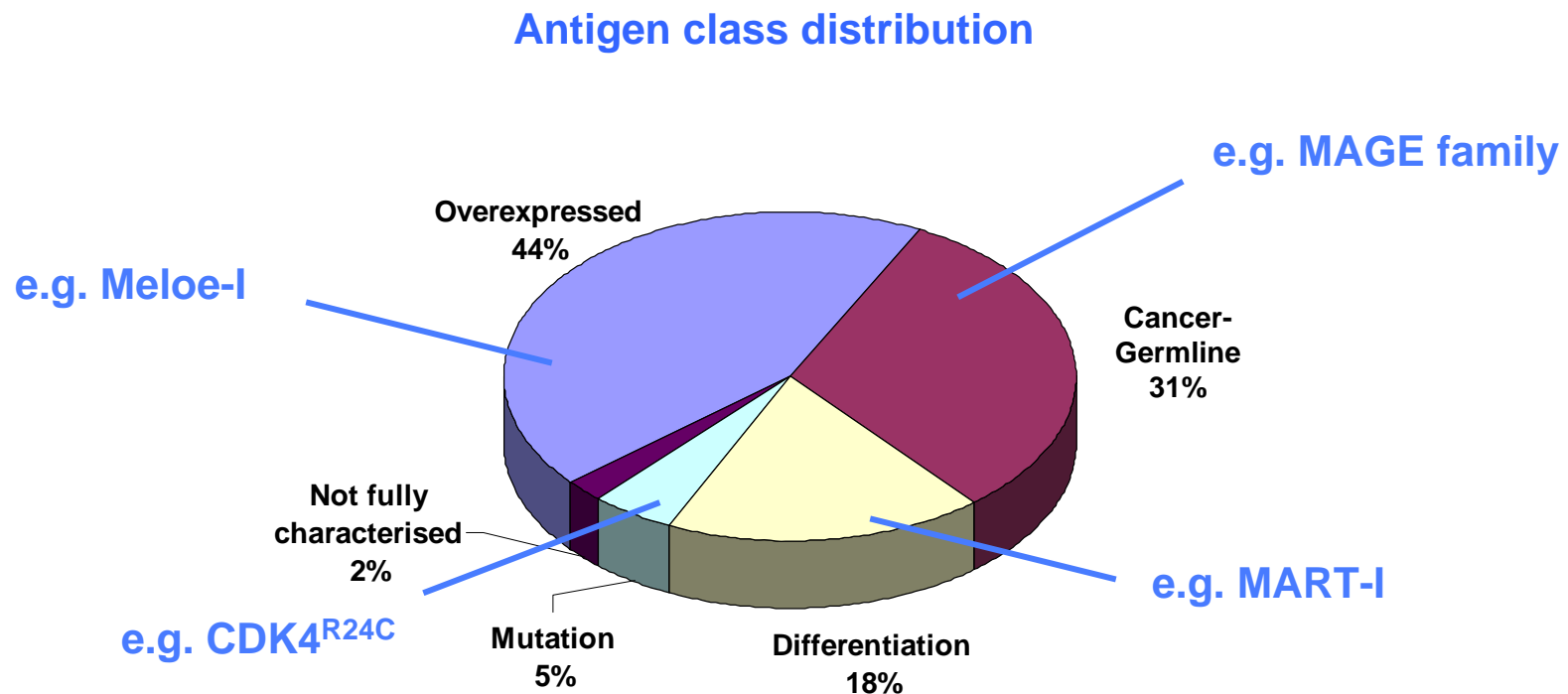


## Selection of melanoma-associated epitope panel

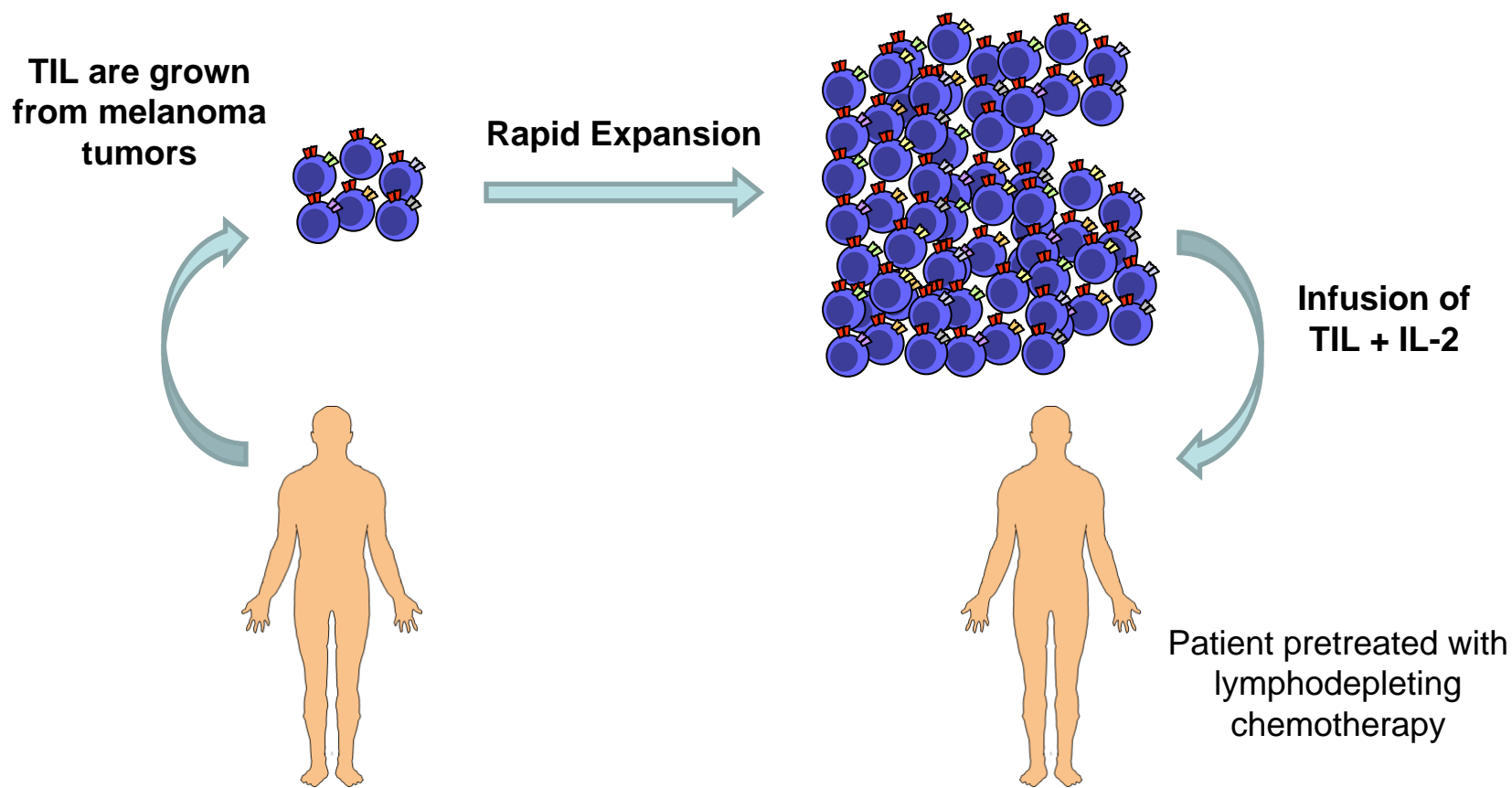
---

### Panel of 216 melanoma-associated peptides

- public databases
- literature analysis



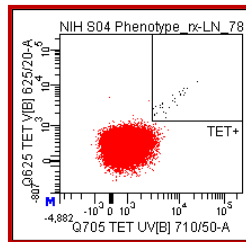
## Dissecting therapy-induced T cell responses in melanoma: POC



- Do we detect tumor-specific T cell responses in the cell product?
  - If so, what do these T cells recognize?
- Does composition of the cell product predict post-treatment immune reactivity?

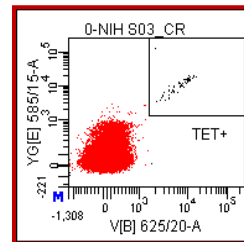
# Identification of diverse T cell populations in HLA-A2+ TIL products

pt.LN



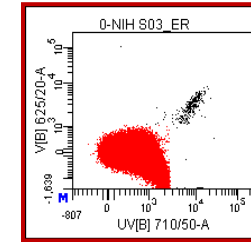
**MART-1 (#78)**  
(0.087%)  
ELAGIGILTV

pt.CR



**MART-1 (#78)**  
(0.113%)  
ELAGIGILTV

pt.ER

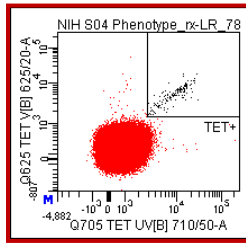


**MART-1 (#78)**  
(0.074%)  
ELAGIGILTV

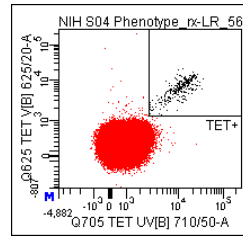
Numbers represent the % MHC tetramer+ T cells out of total CD8+ cells

# Identification of diverse T cell populations in HLA-A2+ TIL products

pt.LR

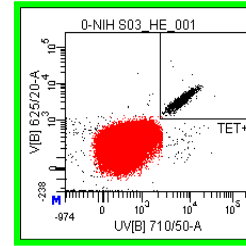


**MART-1 (#78)**  
(0.018%)  
ELAGIGILTV

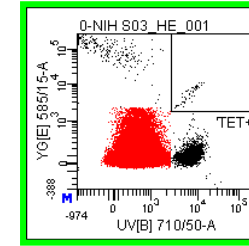


**gp100 (#56)**  
(0.045%)  
IMDQVPFSV

pt.HE

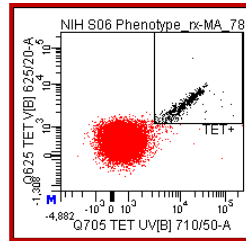


**SSX-2 (#114)**  
(0.628%)  
KASEKIFVY

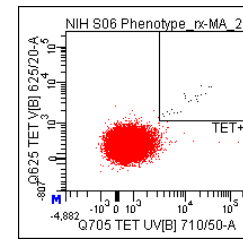


**MAGE-A10 (#59)**  
(0.015%)  
GLYDGM EHL

pt.MA

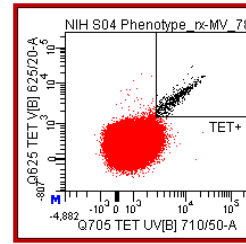


**MART-1 (#78)**  
(4.581%)  
ELAGIGILTV

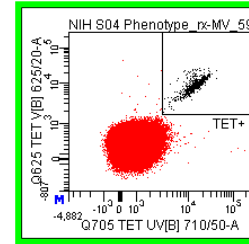


**gp100 (#56)**  
(0.071%)  
IMDQVPFSV

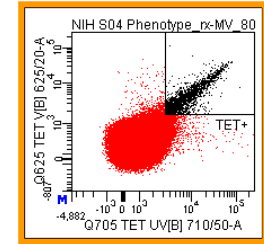
pt.MV



**MART-1 (#78)**  
(0.150%)  
ELAGIGILTV



**MAGE-A10 (#59)**  
(0.137%)  
GLYDGM EHL



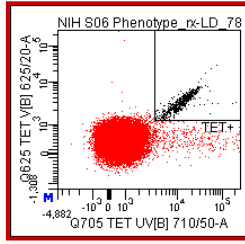
**Meloe-1 (#80)**  
(0.344%)  
TLNDECWPA

Numbers represent the % MHC tetramer+ T cells out of total CD8+ cells

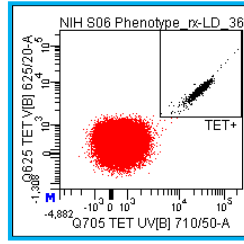


# Identification of diverse T cell populations in HLA-A2+ TIL products

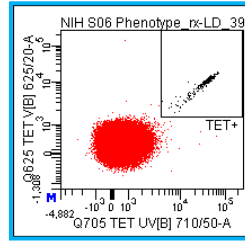
pt.LD



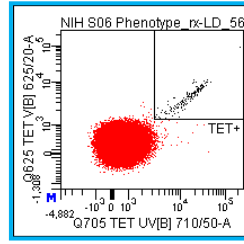
**MART-1 (#78)**  
1.210%  
ELAGIGILTV



**Gp100 (#36)**  
1.897%  
YLEPGPVTA

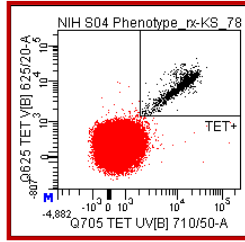


**Gp100 (#39)**  
0.131%  
VLYRYGSFSV

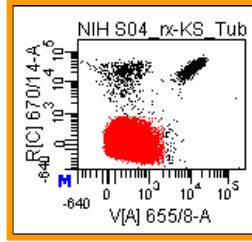


**Gp100 (#56)**  
0.083%  
IMDQVPFSV

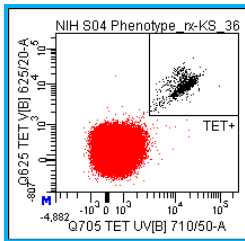
pt.KS



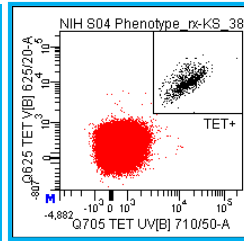
**MART-1 (#78)**  
0.633%  
ELAGIGILTV



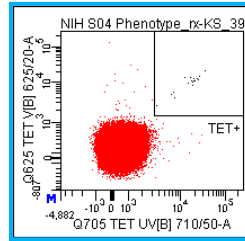
**Meloe-1 (#80)**  
(1.45%)  
TLNDECWPA



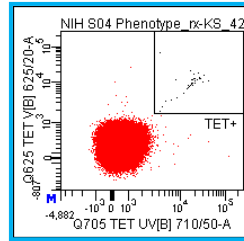
**gp100 (#36)**  
0.453%  
YLEPGPVTA



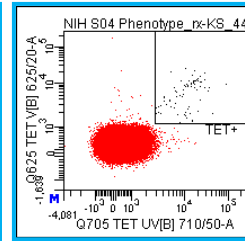
**gp100 (#38)**  
0.376%  
ITDQVPFSV



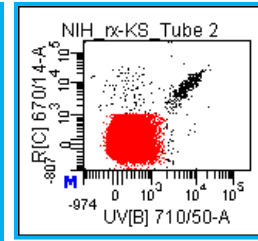
**gp100 (#39)**  
0.010%  
VLYRYGSFSV



**gp100 (#42)**  
0.016%  
AMLGHTTMEV



**gp100 (#44)**  
0.028%  
KTRWGQYWQV



**gp100 (#56)**  
(0.42%)  
IMDQVPFSV



## Summary of identified T cell responses in 15 T cell products

	Epitope	Patient														
		PSC	CZE	RAV	RTE	MZU	LR	LN	KS	MV	CR	ER	MA	LD	SW	HE
Differentiation	Mart-1 <sub>ELA</sub>	Red			Green	Green	Green	Green	Green	Green	Green	Green	Orange	Orange		
	gp100 <sub>IMD</sub>						Green	Green	Green		Green	Green	Green	Green		
	gp100 <sub>YLE</sub>													Orange		
	gp100 <sub>VLY</sub>													Green		
	gp100 <sub>ITD</sub>															
	gp100 <sub>AML</sub>															
	gp100 <sub>KTW</sub>															
Cancer/ Germline	TRP2 <sub>VYD</sub>	Red														
	TRP2 <sub>SVY</sub>	Red														
OE	MAGE A10 <sub>GLY</sub>				Red				Green							Green
	MAGE C2 <sub>LLF</sub>	Orange			Green											
	MAGE C2 <sub>ALK</sub>				Green											
	MAGE C2 <sub>VIW</sub>				Orange											
	SSX-2 <sub>KAS</sub>															Green
M	Meloe-1 <sub>TLN</sub>		Green						Orange	Green						
	Telomerase <sub>RLF</sub>	Green														
	CDK4 <sub>ACD</sub>				Orange											
U	MG50 <sub>RLG</sub>	Green			Green											

- Total of 39 melanoma-reactive T cell responses in 15 T cell products
- Number of responses ranges from 0 to 8
- Predominant recognition of MDA and C/G antigens
- T cell reactivity is *highly* patient-specific

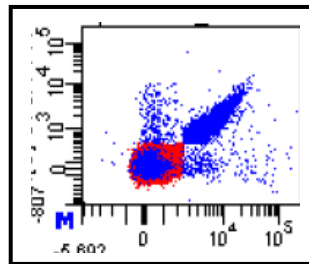
## Summary of identified T cell responses in 15 T cell products

	Epitope	Patient														
		PSC	CZE	RAV	RTE	MZU	LR	LN	KS	MV	CR	ER	MA	LD	SW	HE
Differentiation	Mart-1 <sub>ELA</sub>	Red			Green	Green	Green	Green	Green	Green	Green	Green	Orange	Orange		
	gp100 <sub>IMD</sub>						Green	Green	Green		Green	Green	Green	Green		
	gp100 <sub>YLE</sub>													Orange		
	gp100 <sub>VLY</sub>													Green		
	gp100 <sub>ITD</sub>															
	gp100 <sub>AML</sub>															
	gp100 <sub>KTW</sub>															
Cancer/ Germline	TRP2 <sub>VYD</sub>	Red														
	TRP2 <sub>SVY</sub>	Red														
OE	MAGE A10 <sub>GLY</sub>				Red				Green							Green
	MAGE C2 <sub>LLF</sub>	Orange			Green											
	MAGE C2 <sub>ALK</sub>				Green											
	MAGE C2 <sub>VIW</sub>				Orange											
	SSX-2 <sub>KAS</sub>															Green
	Meloe-1 <sub>TLN</sub>								Orange	Green						
M	Telomerase <sub>RLF</sub>	Green														
	CDK4 <sub>ACD</sub>				Orange											
	MG50 <sub>RLG</sub>	Green			Green											

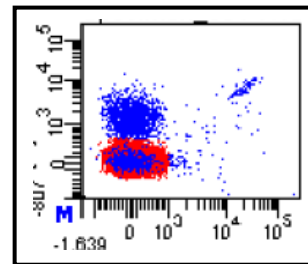
- Total of 39 melanoma-reactive T cell responses in 15 T cell products
- Number of responses ranges from 0 to 8
- Predominant recognition of MDA and C/G antigens
- T cell reactivity is *highly* patient-specific

- Are these T cell populations relevant for melanoma recognition?
- Does this analysis predict post-treatment immune reactivity?

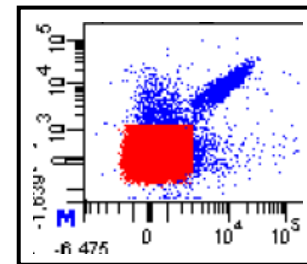
# Isolation of a melanoma-reactive T cell populations for functional validation



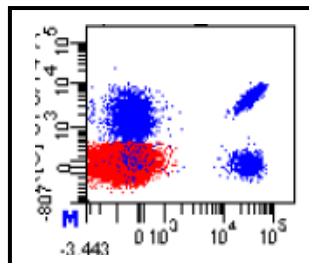
**MAGE-A10**  
5.3%  
GLYDGMEHL



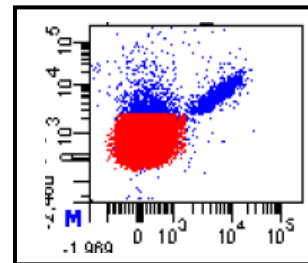
**MAGE-C2**  
0.2%  
ALKDVEERV



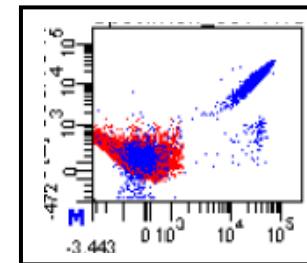
**MAGE-C2**  
0.9%  
LLFGLALIEV



**MAGE-C2**  
1.5%  
VIWEVLNAV



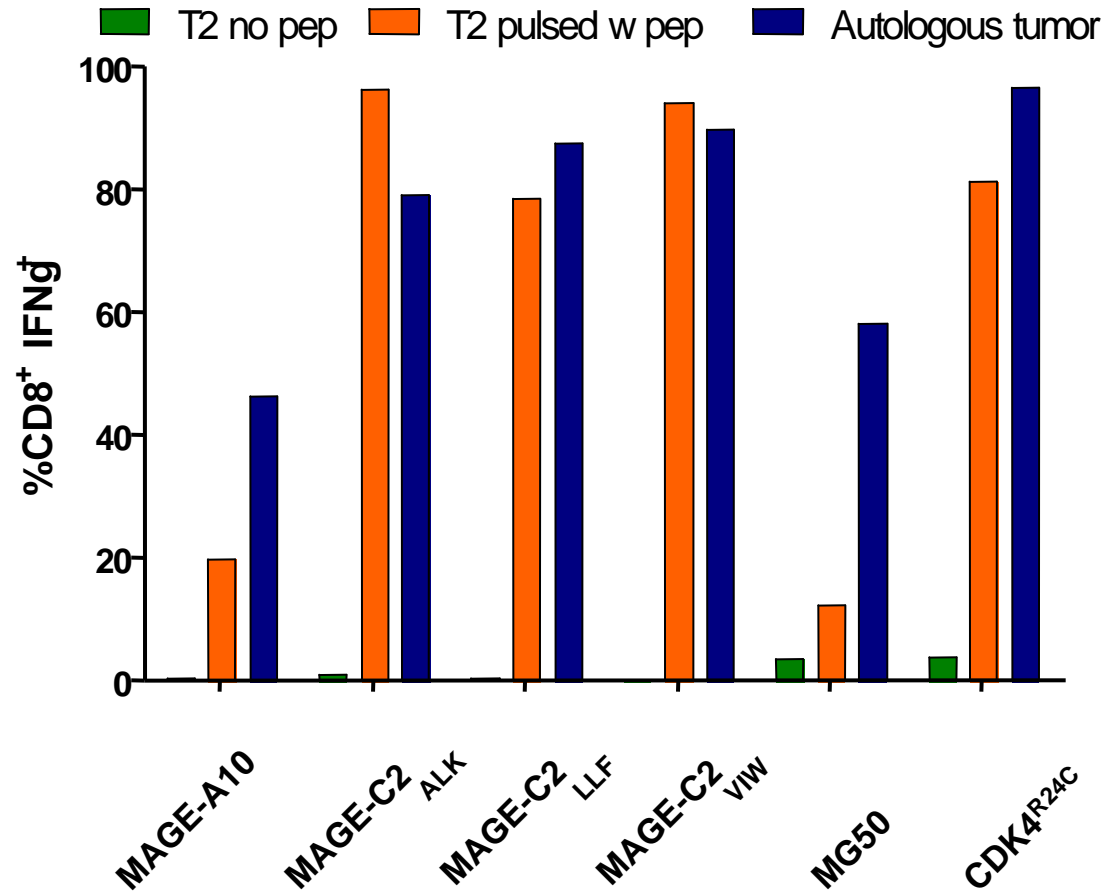
**MG-50**  
0.8%  
RLGPTLMCL



**CDK4 R24C**  
2.3%  
ACDPHSGHFV

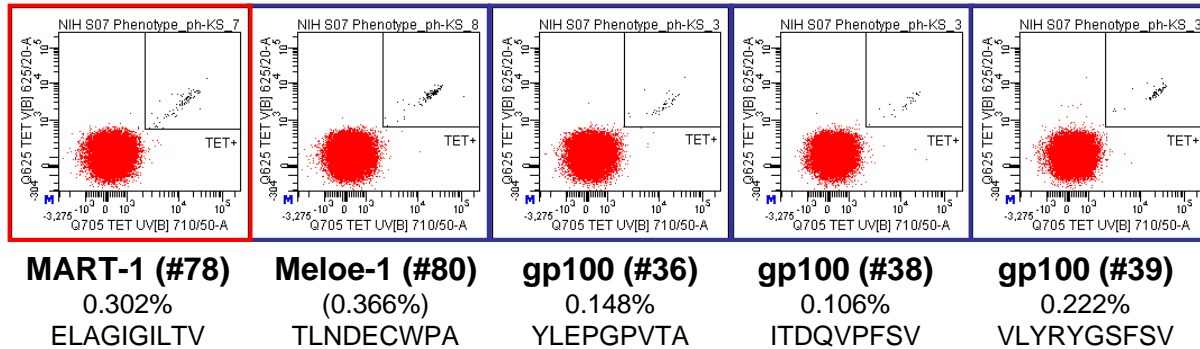
Numbers represent the % MHC tetramer<sup>+</sup> T cells out of total CD8<sup>+</sup> cells

# Recognition of autologous melanoma by identified T cell populations



# Detecting *post-therapy* T cell reactivity in patients treated with TIL

pt.KS  
Post-Infusion PBMNC



- Total of 19 post-infusion responses in 10 HLA-A2<sup>+</sup> TIL-treated patients
- 79% of pre-infusion responses is seen back in post-infusion PBMNC
- 95% of T cell responses in PBMNC post-therapy is predicted by in vitro reactivity

## Conclusions

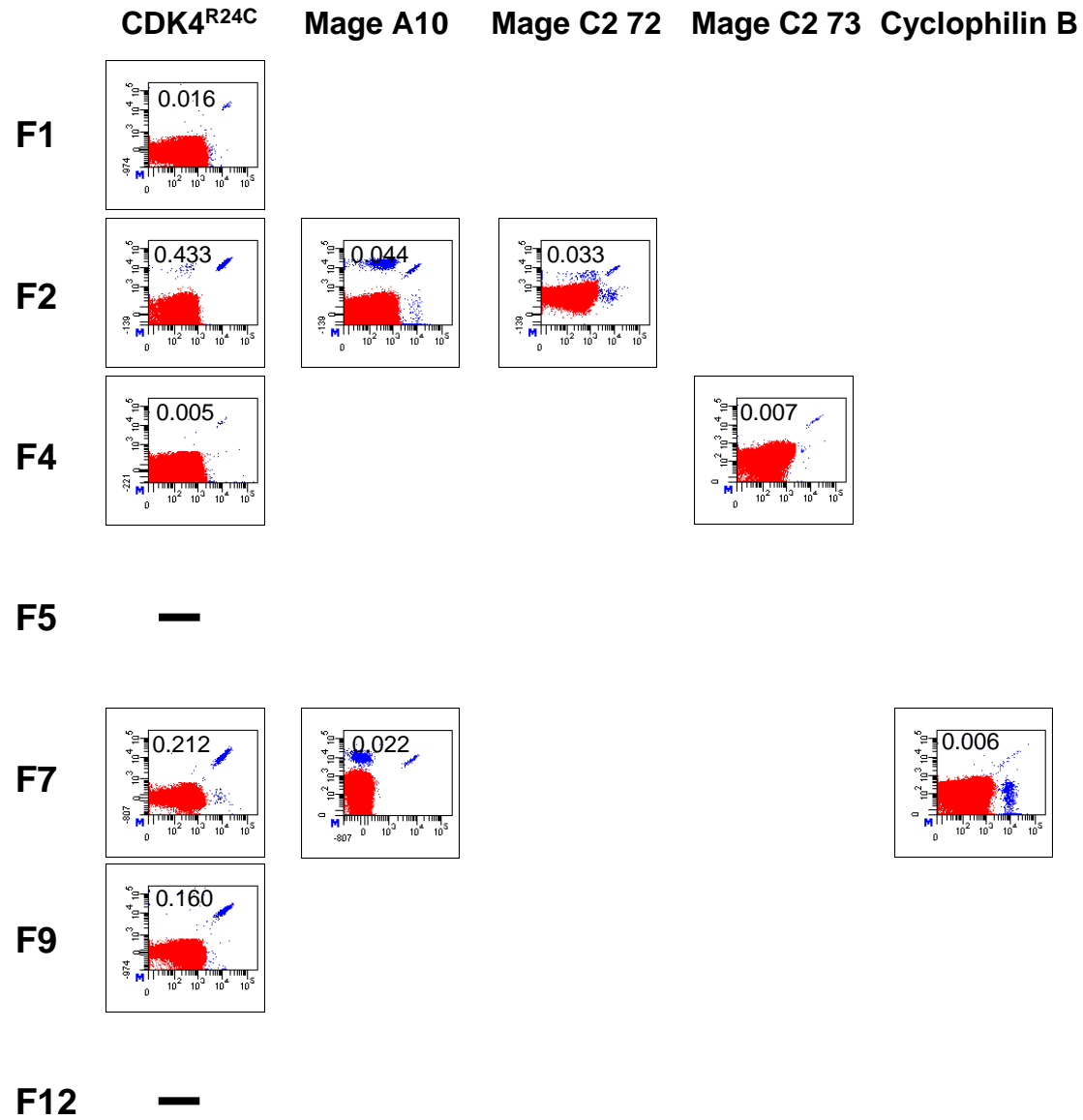
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- ❖ We have developed and validated technology for the high-throughout detection of therapy-induced melanoma-specific T cell reactivity
- ❖ T cell responses in TIL cell products are diverse, biased towards MDA and C/G antigens, and highly variable between patients
- ❖ In 'young TIL', individual T cell responses are generally of a (surprisingly) low magnitude
- ❖ T cell responses in TIL cell products predict immune reactivity post-therapy
  - No epitope spreading -

Is it possible to influence TIL cell product composition?



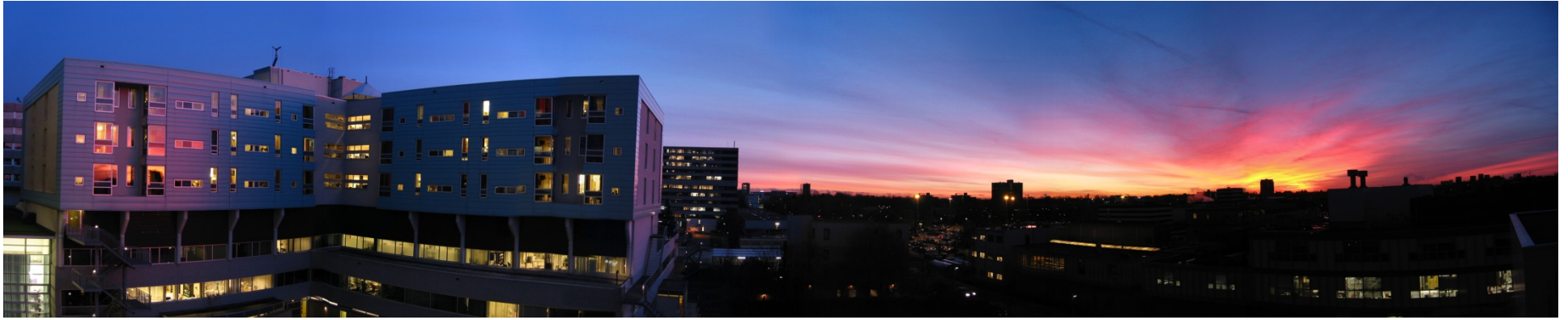
# Individual tumor fragments yield distinct TIL products



## Dissecting melanoma-specific T cell responses upon other immunotherapeutic treatments?

---

- ❖ Immune reactivity induced by anti-CTLA4 or anti-PD1 treatment?
- ❖ Immunological consequences of combination therapy?  
(e.g. BRAF inhibition plus anti-CTLA4)



## Self-destructive MHC ligands & combinatorial coding

### MHC-based monitoring

Jenny Shu

Manuel Fankhauser

Pia Kvistborg

Mireille Toebes

Arne Bakker

Carsten Linnemann

### Chemical Biology

Boris Rodenko

Huib Ovaa

### Clinical translation

Bianca Heemskerk

Annelies Jorritsma

Raquel Gomez

Nienke van Rooij

Bastiaan Nuijen

Christian Blank

John Haanen

### CCIT, Copenhagen

Sine Hadrup

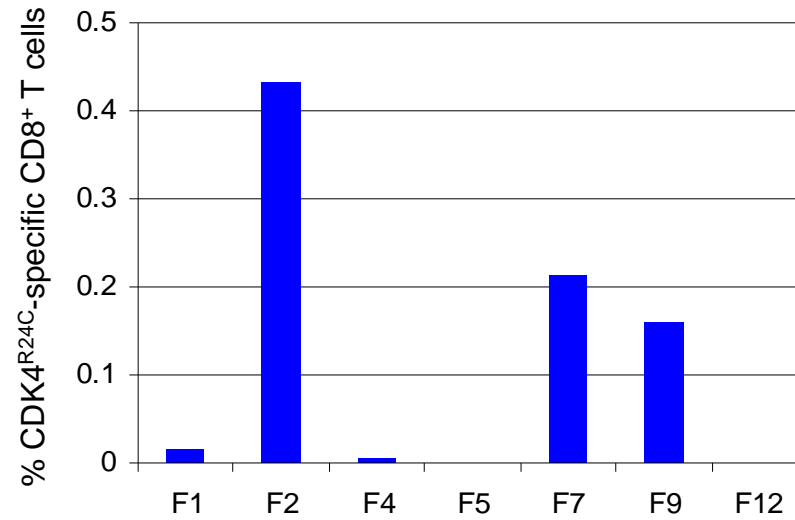
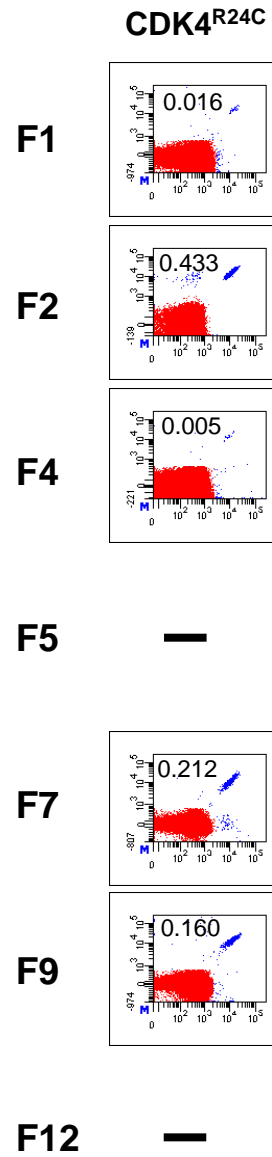
Charlotte Albæk Thruø

### Surgery Branch, NIH

Steven Rosenberg

Mark Dudley

# Individual tumor fragments yield distinct TIL products



# Self-assembling molecular codes

